Households contaminated by environmental tobacco smoke: sources of infant exposures

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Objectives: To examine (1) whether dust and surfaces in households of smokers are contaminated with environmental tobacco smoke (ETS); (2) whether smoking parents can protect their infants by smoking outside and away from the infant; and (3) whether contaminated dust, surfaces, and air contribute to ETS exposure in infants.

Design: Quasi-experiment comparing three types of households with infants: (1) non-smokers who believe they have protected their children from ETS; (2) smokers who believe they have protected their children from ETS; (3) smokers who expose their children to ETS.

Setting: Homes of smokers and non-smokers.

Participants: Smoking and non-smoking mothers and their infants < 1 year.

Main outcome measures: ETS contamination as measured by nicotine in household dust, indoor air, and household surfaces. ETS exposure as measured by cotinine levels in infant urine.

Results: ETS contamination and ETS exposure were 5–7 times higher in households of smokers trying to protect their infants by smoking outdoors than in households of non-smokers. ETS contamination and exposure were 3–8 times higher in households of smokers who exposed their infants to ETS by smoking indoors than in households of smokers trying to protect their children by smoking outdoors.

Conclusions: Dust and surfaces in homes of smokers are contaminated with ETS. Infants of smokers are at risk of ETS exposure in their homes through dust, surfaces, and air. Smoking outside the home and away from the infant reduces but does not completely protect a smoker’s home from ETS contamination and a smoker’s infant from ETS exposure.

Environmental tobacco smoke (ETS)—also known as secondhand smoke—is a complex mixture of more than 4000 chemical compounds that are generated during the burning of tobacco products. This mixture contains numerous irritants and toxicants with acute health effects as well as toxicants with carcinogenic effects in humans. ETS is known to increase morbidity and mortality risks in infants, children, and adult non-smokers.

Data from the California Tobacco Survey (CTS) indicate that, in 1999, 72.8% of homes in California were smoke-free, leaving approximately one in four homes at risk of contributing to tobacco exposure of non-smokers. In homes with children under 6 years of age where all adults smoked, 56.7% of respondents reported having a complete smoking ban. In homes with children where only some adults smoked, 73.1% were reportedly smoke-free in 1999. Similar patterns emerge for the USA at large. Data from the National Health and Nutrition Examination Survey III show that 43% of US children (aged 2 months to 11 years) lived in a home with at least one smoker, and that 37% of adult non-tobacco users lived in a home with a smoker or reported exposure to ETS at work.

More recently, the 2000 Behavioral Risk Factor Surveillance System collected data from 20 states about smoking policies at home. The percentage of adults reporting no smoking at home ranged from 61% (Virginia) to 79% (Colorado), suggesting that 20–40% of US homes contribute to tobacco exposure of non-smokers.

The best understood route of exposure to ETS is the inhalation of contaminated indoor air. In addition to gas and vapour phase ETS components, contaminated air also contains ETS particles. Because the ETS particles have a mass median aerodynamic diameter of well below 2.5 μm, they become respirable suspended particles (RSPs) that cannot be easily filtered and removed by the protective mechanisms of nose and throat. The size of these particles allows them to enter the deep lung and to cause damage due to their size alone. To this effect can be added the chemical toxicity of the particles that enter the deep lung. Thus, both the size of ETS particles and the systemic effects of the chemical toxicity of ETS components may contribute to morbidity.

Inhaling ETS while a cigarette is being smoked is the most noticeable, though not the only exposure occasion. From ETS chamber and field studies, it is known that ETS components are rapidly dispersed after emission and undergo further dynamic chemical reactions. Vapour phase components deposit and are adsorbed onto walls, furniture, clothes, toys, and other objects within 10 of minutes to hours after tobacco smoke has been emitted. From there, they are re-emitted into the air over the course of hours to months. ETS particulate matter can deposit on surfaces within hours after smoking occurred, from where it may be re-suspended or react with vapour phase compounds. Through this dynamic behaviour, ETS can contaminate house dust, carpets, walls, furniture, and other household objects for weeks and months after ETS was emitted from a cigarette. Findings from controlled chamber and field studies suggest that residential indoor...
environments become reservoirs for ETS, turning contaminated dust, carpets, and other household objects into potential sources of ETS exposure long after smoking has stopped.\textsuperscript{1, 10}

Infants of smoking parents are at a particular risk of second-hand smoke exposure through contaminated house dust and surfaces. During their first year of life, infants spend much time indoors, are in close proximity to contaminated dust and objects (for example, blankets, carpets, floors), and are in close physical contact with their smoking parents. At approximately 0.05–0.25 g/day, the dust ingestion rate in infants is estimated to be more than twice that of adults.\textsuperscript{11} Moreover, because of their developmental stage, infants exhibit a much higher frequency of hand-to-mouth and object-to-mouth contacts and ingestion of non-food items (that is, pica behaviour) than older children or adults.\textsuperscript{12} In addition to increased inhalation of contaminated dust, infants may also be exposed to ETS through ingesting and touching contaminated objects and surfaces. As infants and young children are highly active near the floor, they may also be exposed to higher levels of re-suspended floor dust than adults.

House dust and contaminated surfaces are known to be a major source of contaminants such as lead, allergens, pesticides, and polycyclic aromatic hydrocarbons (PAH).\textsuperscript{13} However, little research is available on ETS contamination of house dust. Hein \textit{et al}\textsuperscript{14} were the first to detect nicotine in house dust from homes of smokers. They compared house dust from homes of 34 smokers and 38 non-smokers and found a strong positive correlation ($r = 0.65$) between amount smoked and the nicotine concentration in the house dust. The amount of nicotine inhaled during one hour was estimated for someone in a home with high nicotine concentration in the house dust to be 12 ng, a relatively small amount compared to that inhaled by an active smoker (600–3000 ng/h). However, considering that an infant may spend the entire day indoors, has a higher respiration rate (factor 3–8) and a lower body weight than an adult (factor 10–20), this relatively low dosage of ETS exposure may accumulate over the course of weeks to levels equivalent to several hours of active adult smoking. Thus, long term exposure to contaminated house dust raises the possibility of significant exposure to toxic agents in ETS, which might contribute to disease aetiology or exacerbation of pre-existing illness.

This study explored the potential role of dust and surface contamination as sources of exposure to the contents of ETS for infants. We compared three types of households. The “direct exposure group” (DEG) consisted of households in which parents smoked indoors at home and in the presence of their child. The “indirect exposure group” (IEG) consisted of households in which parents smoked and attempted to protect their infants by smoking outside of the home and in the absence of their child. The control group (“no exposure group”, NEG) consisted of households with parents who have never smoked, in which no smoking took place indoors, and the infant was not knowingly exposed to tobacco smoke elsewhere. Multiple measures of air, dust, and surface contamination and multiple measures of the infants’ exposure to tobacco smoke were examined to address the following questions:

- Are house dust and surfaces in households of smokers contaminated with secondhand smoke?
- Do smoking parents protect their infants by smoking outside and away from the infant?
- Do contaminated household dust and surfaces contribute to the overall exposure of infants to secondhand smoke?

\section*{METHODS}

\subsection*{Participants}
Participants were recruited through advertisement at WIC (Women, Infants, and Children Supplemental Food and Nutrition Program; 96\%) sites in San Diego County and in the local news media (4\%). Interested mothers were contacted by phone to determine their eligibility. To qualify, all mothers had to have an infant between 2–12 months old and could not have breast fed their baby within the past 30 days. Though we understand that breast feeding may enhance the health of an infant, we elected to omit families where the mother was breastfeeding the infant because breast feeding by a smoker (or mother exposed to ETS) may transmit nicotine and confound our cotinine measures of ETS. Subjects were paid up to $100 for participating in the study. Forty nine infants aged 2–13 months and their mothers completed the study.

Table 1 provides sociodemographic information about the household, mothers, and infants in the three exposure groups. The three groups did not differ (all $p > 0.15$) with respect to household size and income, size of the home, age and sex of the infant, and mother’s age and employment status. Mothers in the no-exposure group tended to be more educated (35\% completed college) than mothers in the indirect (6\%) and direct (0\%) exposure groups ($\chi^2(2) = 11.0, p = 0.004$). Moreover, the proportion of white non-Hispanic mothers was lower in the IEG (41\%) than the IEG (69\%) and DEG (75\%) households, although this difference was not significant ($\chi^2(2) = 4.5, p = 0.103$).

\subsection*{Design and setting}
This study relied on a non-equivalent group design, comparing three types of households in which infants were not exposed, indirectly exposed, or directly exposed to tobacco smoke. To qualify for the no exposure control group (NEG), all of the following conditions had to be met at the time of screening: (1) all household residents were non-smokers (that is, consumed no tobacco products) for at least one year; (2) no regular visitors smoked in the residence during the last year; (3) no visitors (regular or occasional) smoked cigarettes in the residence within the past 30 days; (4) there were no visits to a home where someone smoked in the same room with the infant within the past 30 days. In summary, NEG households ($n = 17$) serve as the baseline for ETS contamination and exposure measures.

To qualify for the indirect exposure group (IEG), all of the following conditions had to be met: (1) the mother had to smoke every day and at least 20 cigarettes/week over the past three months; (2) the mother or other household residents may not have smoked any cigarettes in the same room (or car) with the infant over the past three months; (3) the mother must have smoked at least 10 cigarettes/week at home outside or in a different room from the infant over the past three months. To rule out that smoking indoors at home in a different room contributed to direct ETS exposure, we identified households in which reportedly no indoor smoking took place during the assessment period. Findings are reported separately for all IEG households and those without indoor smoking. In summary, IEG households ($n = 17$) represent smoking parents who have made serious attempts to protect their children from ETS by not smoking in their presence. This group comes closest to what are commonly referred to as households with smoking bans.\textsuperscript{1}

To qualify for the direct exposure group (DEG) all of the following conditions had to be met: (1) the mother had to smoke every day and at least 20 cigarettes/week over the past three months; (2) the mother or other household residents must have smoked at least 20 cigarettes per week at home
over the past three months; (3) the mother or other household residents had to smoke one or more cigarettes per day (or seven cigarettes per week) at home in the same household; (4) the average number of cigarettes per day smoked indoors by the mother or other household residents; and (5) the mother’s average number of cigarettes smoked per day.

### Measures

#### Measurement schedule

Each residence was visited three times over the course of one week. All visits took place on a Tuesday, Friday, Monday schedule or a Friday, Monday, and Thursday schedule, such that all measures included exposures during a weekend. Each visit consisted of an interview with the mother, the collection of dust and surface wipe samples in the living room and the sleeping area. Areas were carefully measured from reference points, and all measures included exposures during a weekend. All diary based measures reflect behaviours over the seven study days only. The interview (1), (2), and (3) and diary based measures (5), (6), and (7) assessed the same behaviours over slightly different reference periods. Thus, they served as a check for consistency between retrospective reported behaviour and prospectively recorded practices, but they could not be compared directly because they represented slightly different time frames.

### Air nicotine in living room and bedroom

Air levels of vapour phase nicotine were measured with passive diffusion monitor badges developed by Hammond et al. and used by us previously. The badges were placed in the baby’s home for the duration of the week, placed on the first and removed on the third visit. One badge was placed in the living area and one in the baby’s sleeping area. The height of the monitors was 2 feet from the floor, and badges were placed away from doors and windows. Unmarked non-analysed badges were placed in all other rooms such that all rooms appeared to have air monitors, in keeping with a bogus pipeline procedure. This was employed to prevent smokers moving to a room without a monitoring badge. The number of hours placed in the home was recorded. The badges consisted of a modified 37 mm diffusive sampling cassette with a sodium bisulfate treated Teflon coated glass fibre filter. The badges were stored at −20°C and sent to K Hammond (University of California, Berkeley) for analysis as previously described. Briefly, the nicotine was extracted into hexane and analysed on a gas chromatograph with a nitrogen detector, and results expressed as μg of nicotine/m³ of air. The level of detection for one full week of exposure is 0.02 μg/m³.

### Dust nicotine in living room and bedroom

Two area floor dust samples per visit were collected with a high volume, small surface sampler (HS53, CS-3 Inc, Sandpoint, Idaho, USA), from a 150 cm × 150 cm area, if possible. Some homes had a smaller area sampled, with none being less than 100 cm × 100 cm. One sample was obtained from the living room area and the other from the infant’s sleeping area. Areas were carefully measured from reference points in the home to allow collection of dust from the same area each time, without leaving any marks visible to the occupants. Floor dust samples were collected into Teflon

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**Table 1** Demographic characteristics of participants in different exposure groups

<table>
<thead>
<tr>
<th>Demographic characteristics</th>
<th>No exposure</th>
<th>Indirect exposure</th>
<th>Direct exposure</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sample size</td>
<td>17</td>
<td>17</td>
<td>15</td>
</tr>
<tr>
<td>Infant</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Female (%)</td>
<td>59</td>
<td>56</td>
<td>47</td>
</tr>
<tr>
<td>Mean age (months)</td>
<td>7.6</td>
<td>6.1</td>
<td>7.7</td>
</tr>
<tr>
<td>Race/ethnicity (%)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>African American</td>
<td>18</td>
<td>6</td>
<td>19</td>
</tr>
<tr>
<td>White non-Hispanic</td>
<td>29</td>
<td>69</td>
<td>63</td>
</tr>
<tr>
<td>Mexican American</td>
<td>12</td>
<td>0</td>
<td>6</td>
</tr>
<tr>
<td>Multiracial</td>
<td>29</td>
<td>0</td>
<td>13</td>
</tr>
<tr>
<td>Other</td>
<td>6</td>
<td>19</td>
<td>0</td>
</tr>
<tr>
<td>Mother</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Race/ethnicity (%)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>African American</td>
<td>18</td>
<td>6</td>
<td>13</td>
</tr>
<tr>
<td>White non-Hispanic</td>
<td>41</td>
<td>69</td>
<td>75</td>
</tr>
<tr>
<td>Mexican American</td>
<td>18</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Multiracial</td>
<td>12</td>
<td>0</td>
<td>6</td>
</tr>
<tr>
<td>Other</td>
<td>12</td>
<td>25</td>
<td>6</td>
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<tr>
<td>Education level</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Did not complete high school</td>
<td>18</td>
<td>0</td>
<td>13</td>
</tr>
<tr>
<td>Completed high school</td>
<td>12</td>
<td>44</td>
<td>44</td>
</tr>
<tr>
<td>Technical/vocational school</td>
<td>0</td>
<td>6</td>
<td>25</td>
</tr>
<tr>
<td>Some college</td>
<td>41</td>
<td>44</td>
<td>19</td>
</tr>
<tr>
<td>Completed college</td>
<td>35</td>
<td>6</td>
<td>0</td>
</tr>
<tr>
<td>Employment status</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Not employed</td>
<td>71</td>
<td>81</td>
<td>56</td>
</tr>
<tr>
<td>Part time (&lt;40 hours)</td>
<td>12</td>
<td>6</td>
<td>25</td>
</tr>
<tr>
<td>Full-time (&gt;40 hours)</td>
<td>19</td>
<td>13</td>
<td>19</td>
</tr>
<tr>
<td>Household</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total income (median)</td>
<td>$24000</td>
<td>$21000</td>
<td>$25000</td>
</tr>
<tr>
<td>Size of residence (median, square feet)</td>
<td>639</td>
<td>586</td>
<td>736</td>
</tr>
<tr>
<td>Number residents (median)</td>
<td>4</td>
<td>4.5</td>
<td>4</td>
</tr>
</tbody>
</table>

*p < 0.05; all other group differences are not statistically significant (p > 0.10).*
bottles, transported cooled, weighed, and sieved with a stainless steel, methanol washed, 150 μm mesh sieve. Sieved dust was weighed and stored in glass bottles at −70°C until analysis. Analysis for nicotine was performed on a gas chromatograph equipped with mass spectrometry (GC-MS, HP 6890) using a method adapted from one developed at the US Centers for Disease Control and Prevention (CDC) for analysis of nicotine in wipe samples. Cotinine and its labelled reference methyl-d3 cotinine were originally included in all analysis, but when approximately half of the samples had been analysed with cotinine detected in only two, cotinine was dropped from further analysis. The limit of detection was 0.03 μg/mg dust (CDC method) to 0.002 μg/mg dust (SDSU method, J Polansky modifications).

Surface nicotine on furniture in living room and bedroom

Two area wipe samples per visit were taken with a pre-screened wipe, covering a 10 × 10 cm area. Wipes were soaked in freshly prepared 0.1% (w/v) ascorbic acid to preserve the nicotine. One wipe was taken from the living room area (typically the coffee table). The other wipe was taken from the baby's sleeping area (typically the bed frame). The same locations were wiped each visit. Wipes were placed into glass bottles, transported cooled, and stored at −70°C until analysis. Levels were expressed as weight of nicotine per wipe. The limit of detection was 0.06 μg/wipe (CDC) to 0.01 μg/wipe (SDSU method, J Polansky modifications).

Nicotine on mother’s index finger

A wipe sample of the mother’s index finger from the hand used to hold a cigarette was taken at each visit. Wipe was moistened and processed as above. In order to keep costs down while investigating the hypothesis that some nicotine might be present on mother’s hands, only one sample was chosen for analysis from four mothers in the IEG and four mothers in the DEG groups.

Urine cotinine

Urine samples were collected from the infant at each visit using a standard urine collection bag for infants or a cotton roll placed in the diaper. Cotton rolls were placed in a sterile 20 ml syringe and expressed into sterile 5 ml plastic cryovials. Samples were immediately frozen at −20°C before they were packed in dry ice and shipped to the CDC for analysis using high performance liquid chromatography, atmospheric pressure chemical ionisation tandem mass spectrometry (HPLC APCI-MS³⁰). Cotinine levels reported are “total” cotinine, combining bound and unbound quantities of the metabolite. The assay is sensitive to levels as low as 0.05 ng/ml.

Hair nicotine and cotinine

In combination with the urine cotinine measure (1–3 days’ half life), hair cotinine provides a measure of exposure over a longer period of time (1–2 months). Hair samples were obtained at the last visit by cutting 1 cm of 10–15 hair shafts (approximately 10 mg in weight) close to the scalp from the back of the head (posterior vertex, occipital bone) using methanol cleaned scissors. Samples were stored in sterile vials and sent to J Klein (University of Toronto) for analysis as described.⁵ The limits of detection were 0.02 ng/mg and 0.05 ng/mg for cotinine and nicotine, respectively.

Statistical analyses

Statistical analyses were conducted using STATATA 7.0⁴ and SPSS 10.1. All outcome measures were subjected to logarithmic transformation before analyses were conducted to deal with skewed error distribution and to stabilise error variances. Relations between measures of contamination and exposure were examined using rank order and Pearson product moment (PPM) correlations. Because findings do not differ substantially, we only report those for PPM correlations. Significance was set at α = 0.05.

Differences in outcome measures between groups were tested via Tobit regression models, in which an observation was defined as left censored if the value fell below the detection limit of a particular outcome measure. In addition, we used robust estimates of standard errors based on the Huber-White sandwich estimator of variance⁶ to protect against the undue influence of outliers on statistical tests in this relatively small sample.

The contribution of house dust and surface contamination to overall exposure was examined using OLS regression models with robust standard errors based on the Huber-White sandwich estimator of variance.⁶⁷

RESULTS

Smoking behaviour and smoking policies

Table 2 presents descriptive information regarding smoking behaviour and smoking policies in the three exposure groups. In NEG households, nobody was a smoker and no smokers had reportedly visited during the 30 days before the interview.

The IEG and DEG households did not differ significantly with respect to the number of smokers and the percentage of visitors smoking outside of the home. Mothers in DEG households smoked more than mothers in IEG households based on interview data (9.34 v 5.38 cigs/day; t(30) = 2.49, p = 0.018) but not based on diary data (6.20 v 5.41 cigs/day; t(31) = 0.44, p = 0.662). Moreover, DEG households were more likely than IEG households to have visitors who smoked indoors (66.7% v 6.3%; χ²(1) = 10.2, p < 0.01) during the past 30 days. This difference is also reflected in home policies about smoking. Significantly larger proportions of IEG households declared that smokers at home always or almost always smoked outside (88% v 27%; χ²(2) = 9.3, p < 0.01) and shut the doors or windows when smoking outside (69% v 13%; χ²(2) = 13.6, p < 0.01).

In the IEG households, all mothers were smokers and about two out of three households had one or more additional smokers. Four of the 17 IEG households reported that cigarettes were smoked in the home, for an average of 1.06 cigs/day in these four households. Three of these four households also reported that their infants were in a room or car where cigarettes were smoked at home or away from home. In the three households where this occurred, the infants were directly exposed to an average of 0.38 cigs/day.

To control for the occasional indoor exposure of some infants in the IEG group, we identified a subgroup of IEG households in which reportedly no cigarettes were smoked in the home during the assessment week and infants were not knowingly exposed to tobacco smoke (for example, at home, in a car, at someone else’s home). This was done to investigate whether smoking indoors during the assessment period contributed to ETS contamination at home and the child’s exposure. There were no statistically (all p > 0.20) or practically significant differences on any of the exposure measures between the “no indoor smoking/no direct exposure” IEG subgroup (n = 12) and the “occasional indoor smoking/occasional direct exposure” IEG subgroup (n = 4). Similarly, there were no significant differences between the two IEG subgroups in contamination measures, with the exception of the maximum nicotine loading in living room and bedroom dust (table 3). We also investigated whether excluding the four households with occasional indoor smoking and direct exposure would alter findings concerning group differences between NEG, IEG, and DEG households.
However, this was not the case. Therefore, all subsequent statistical analyses rely on the entire group of 17 IEG households to maintain sufficient statistical power. In tables 2, 3, and 4, we report separately findings for all 17 IEG households and the subgroup of 12 households without indoor smoking.

**Contamination of the indoor home environment**

Table 3 presents the nicotine levels found in the air, in dust, on surfaces, and on fingers in the three exposure groups. To investigate whether contamination levels differed between exposure groups, NEG households were compared to IEG households (contrast 1, C1) and IEG households were compared to DEG households (contrast 2, C2).

**Air nicotine levels**

Nicotine was detected in the living room air and the bedroom air in all smoker households and 97% of non-smoker households. Air nicotine concentrations in the living rooms and infant bedrooms of IEG households were approximately twice those of NEG and DEG households. The geometric mean nicotine concentration in living room air of IEG households was 2.57 μg/m³ (95% CI 1.61 to 3.89), while for DEG households it was 0.32 μg/m³ (95% CI 0.08 to 0.62), and for NEG households it was 0.10 μg/m³ (95% CI 0.06 to 0.15).

**Table 2 Smoking behaviours in different exposure groups**

<table>
<thead>
<tr>
<th>Reported smoking behaviour and home policies</th>
<th>No exposure</th>
<th>Indirect exposure</th>
<th>Direct exposure</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>All</td>
<td>No indoor smoking</td>
<td>Indoor smoking</td>
</tr>
<tr>
<td>Sample size</td>
<td>17</td>
<td>17</td>
<td>13</td>
</tr>
<tr>
<td>Average number of smokers in household</td>
<td>0</td>
<td>1.69</td>
<td>1.75</td>
</tr>
<tr>
<td>Households (%) with</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0 smokers</td>
<td>100</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>1 smoker</td>
<td>0</td>
<td>43.8</td>
<td>41.7</td>
</tr>
<tr>
<td>2 smokers</td>
<td>0</td>
<td>43.8</td>
<td>41.7</td>
</tr>
<tr>
<td>3 or more smokers</td>
<td>0</td>
<td>12.5</td>
<td>16.7</td>
</tr>
<tr>
<td>Households (%) in which</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mother smoked</td>
<td>0</td>
<td>100</td>
<td>100</td>
</tr>
<tr>
<td>Visitors smoked in home in past 30 days</td>
<td>0</td>
<td>93.7</td>
<td>33.3</td>
</tr>
<tr>
<td>Households (%) with doors/windows shut</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Always</td>
<td>81.3</td>
<td>91.7</td>
<td>6.7</td>
</tr>
<tr>
<td>Almost always</td>
<td>6.3</td>
<td>0</td>
<td>20.0</td>
</tr>
<tr>
<td>Often, sometimes</td>
<td>12.5</td>
<td>8.3</td>
<td>53.3</td>
</tr>
<tr>
<td>Rarely, never</td>
<td>0</td>
<td>0</td>
<td>20.0</td>
</tr>
<tr>
<td>Households (%) with smokers go outside</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Always</td>
<td>50.0</td>
<td>66.7</td>
<td>6.7</td>
</tr>
<tr>
<td>Almost always</td>
<td>18.8</td>
<td>16.7</td>
<td>6.7</td>
</tr>
<tr>
<td>Often, sometimes</td>
<td>12.5</td>
<td>8.3</td>
<td>20.0</td>
</tr>
<tr>
<td>Rarely, never</td>
<td>18.8</td>
<td>8.3</td>
<td>66.7</td>
</tr>
<tr>
<td>Smoking (geometric mean, 95% CI)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mother’s average cigs/day (diary 7 days)</td>
<td>0</td>
<td>5.41 (3.33 to 9.49)</td>
<td>5.20 (2.72 to 9.35)</td>
</tr>
<tr>
<td>Mother’s average cigs/day (interview past 10 days)</td>
<td>0</td>
<td>5.38 (3.40 to 8.25)</td>
<td>4.44 (2.45 to 9.35)</td>
</tr>
</tbody>
</table>

CI, confidence interval.
three and two times higher, respectively, than those found in the living and bedroom of NEG households (0.32 μg/m² v 0.10 μg/m²; 0.22 μg/m² v 0.09 μg/m²). Air nicotine levels in living rooms and infant bedrooms of DEG households were eight times and seven times higher than in IEG households (2.57 μg/m² v 0.32 μg/m²; 1.50 μg/m² v 0.22 μg/m²).

Significant differences were found in air nicotine levels between NEG and IEG (C1) and between IEG and DEG (C2) in the living room (that is, \( \chi^2(2) = 38.75, p < 0.001; \) C1: t(46) = 6.19, p < 0.001; C2: t(46) = 9.69, p < 0.001) and bedrooms (that is, \( \chi^2(2) = 38.75, p < 0.001; \) C1: t(46) = 4.77, p < 0.001; C2: t(46) = 7.62, p < 0.001). These findings suggest that while parents in the IEG were able to reduce air nicotine levels compared to DEG households, their children were not protected from exposure to nicotine in the indoor air at home.

### Surface nicotine levels

No nicotine was detected on surfaces examined in the living room and infant bedrooms of NEG households. In IEG households, 51% and 53% revealed nicotine levels above the limit of detection on living room and bedroom surfaces, respectively. The average level of the highest nicotine level per household was 19.89 μg/m²; the average of the mean nicotine level per household was 10.68 μg/m².

Nicotine was detected on 88% (49 of 56 samples) of the living room and 88% (35 of 40 samples) of bedroom surfaces in DEG households. Nicotine contamination of surfaces in DEG households was three to five times higher than those found in IEG households. On living room and bedroom surfaces, the average of the highest nicotine levels per household were 73.05 μg/m² and 56.26 μg/m², respectively. The average levels of the mean nicotine level per household were 51.33 μg/m² and 41.85 μg/m² in the living room and bedroom, respectively.

Tobit regression analyses showed significant differences between IEG and DEG households for surface nicotine levels in the living rooms (that is, \( \chi^2(1) = 9.50, p = 0.002; \) C2: t(30) = 3.21, p = 0.003) and bedrooms (that is, \( \chi^2(1) = 16.29, p < 0.001; \) C2: t(30) = 4.59, p < 0.001). These findings suggest that IEG households had lower nicotine levels on household surfaces compared to DEG households. However, IEG households showed surface contamination significantly higher than zero (see confidence interval in table 3). Wipe samples collected in NEG households revealed no detectable levels of nicotine and had to be excluded from the analyses.

### Nicotine on fingers

No nicotine was detected on the fingers of mothers in the NEG households. However, nicotine was detected on the fingers of 100% and 92% of mothers in the IEG and DEG households, respectively. The average nicotine levels in both groups were 0.63 μg/wipe and 0.65 μg/wipe in the IEG and DEG, respectively. Given the surface area of a typical index finger (< 100 cm²), the average nicotine loading on the fingers of the smoking mothers in the IEG and DEG households is more than twice as high as the nicotine loading on living room surfaces of DEG households. Note that the confidence intervals are noticeably large because of the small sample sizes.

Tobit regression analyses revealed that nicotine levels found on the index fingers of smoking mothers were significantly larger than zero (t(19) = 2.63, p = 0.025). Controlling for smoking frequency, no significant differences were found in finger nicotine between mothers in the IEG and DEG groups (that is, \( \chi^2(2) = 0.06, p = 0.97; \) C2: t(10) = 0.13, p = 0.90).

### Nicotine in household dust

Approximately equal amounts of dust were found in bedrooms and living rooms of IEG and DEG households. On day 1 of dust collection, 1.50 g (95% CI 0.75 to 2.58 g) and 1.21 g (95% CI 0.61 to 2.04 g) were collected in the living rooms and bedrooms of IEG households, and 1.50 g, (95% CI 0.57 to 2.98) and 1.57 g (95% CI 0.72 to 2.82) in the DEG households. Summed across all three dust collections, 3.83 g (95% CI 1.82 to 7.28 g) and 2.42 g (95% CI 1.07 to 4.67 g) were collected in the living rooms and bedrooms of IEG households and 3.07 g (95% CI 1.34 to 6.09 g) and 2.87 g (95% CI 1.50 to 4.99 g) in the DEG households.

Nicotine was detected in 38% and 52% of dust samples taken from the living rooms and bedrooms of IEG households. The averages of highest nicotine levels found in the living rooms and bedrooms of each household were 4.43 μg/m² and 3.22 μg/m², respectively. The averages of the mean nicotine levels per household were 2.22 μg/m² and 0.89 μg/m² for the living rooms and infant bedrooms, respectively.

Nicotine was detected in 55% and 70% of dust samples taken from the living rooms and bedrooms of DEG households.
housesholds. The averages of highest nicotine levels found in the living rooms and bedrooms of each household were 64.0 µg/m² and 15.8 µg/m², respectively. The averages of the mean nicotine levels per household were 6.85 µg/m² and 5.37 µg/m² for the living rooms and infant bedrooms, respectively.

Tobit regression analyses revealed significant differences between dust nicotine levels in the living rooms (that is, \( \chi^2(1) = 5.37, p = 0.02 \); C2: \( t(27) = 2.17, p = 0.04 \)) and bedrooms of IEG and DEG households (that is, \( \chi^2(1) = 5.48, p \approx 0.02 \); C2: \( t(27) = 2.29, p = 0.03 \)). These findings suggest that IEG households had lower dust nicotine levels compared to DEG households. Note that dust samples were analysed from IEG and DEG households only, because pilot data revealed no detectable nicotine levels in nonsmoking household.

**Infant exposure to tobacco**

**Mother reported exposure**

Mothers in the NEG households reported that their infants were not exposed to tobacco smoke either at home or away from home. In the IEG group, 76% of mothers indicated their child was not exposed to tobacco smoke, and 24% reported exposure to tobacco smoke away outside of the home (for example, car, friend’s home). All mothers in the DEG group reported that their child was exposed to tobacco at home as well as away from home. As indicated by the number of cigarettes smoked in the presence of the child per day, infants in IEG households were directly exposed to 0.03 and 0.06 cigs/day according to the interview and behavioural diary, respectively. Infants in the DEG households were directly exposed to 5.57 and 5.75 cigs/day based on interview and diary reports, respectively.

Tobit regression models indicated that mother reported exposure levels in the IEG group were not significantly larger than zero (\( t(28) = 1.75, p = 0.091 \)), indicating that mothers noticed little if any ETS exposure. Infant exposure as reported by mothers differed significantly between IEG and DEG households (that is, \( \chi^2(1) = 14.18, p < 0.001 \); C2: \( t(28) = 3.79, p = 0.001 \)), indicating that smoking in the presence of the child was substantially higher in DEG than in IEG households.

**Urine cotinine**

In the NEG households, infant urine cotinine levels averaged 0.33 ng/ml and 0.43 ng/ml based on the mean and the maximum over the three sample days. Urine cotinine levels of infants in the IEG households were approximately eight times higher based on the average (2.47 ng/ml) and the maximum (3.49 ng/ml) over the three sample days. Compared to the IEG households, urine cotinine levels in the DEG households were more than six times higher. The mean levels were 15.47 ng/ml and 20.43 ng/ml based on the average and the maximum across the three sample days, respectively.

Tobit regression analyses showed significant differences in infant urine cotinine levels between NEG and IEG (C1) and between IEG and DEG (C2) (that is, \( \chi^2(2) = 76.22, p < 0.001 \); C1: \( t(45) = 10.85, p < 0.001 \); C2: \( t(45) = 12.76, p < 0.001 \)). Moreover, urine cotinine levels in the IEG differed significantly from zero (\( t(45) = 19.09, p < 0.001 \)). These findings suggest that while infants in the IEG households showed lower exposure levels compared to DEG households, they were not completely protected from secondhand smoke exposure.

**Hair nicotine and cotinine**

We observed a correlation of \( r = 0.81 \) (\( t(34) = 66.9, p < 0.001 \)) between log transformed nicotine and cotinine levels in hair. Hair nicotine and cotinine levels among children in the NEG households were .53 ng/mg and 0.08 ng/mg, respectively. In comparison, hair nicotine and cotinine levels of infants in the IEG households were more than five times higher at 2.65 ng/mg and 0.52 ng/mg, respectively. Infants in the DEG households showed nicotine and cotinine levels approximately twice as high as those in the IEG households at 5.95 ng/mg and 1.05 ng/mg.

Tobit regression analyses revealed significant differences in infant hair cotinine levels between NEG and IEG (C1) and between IEG and DEG (C2) (that is, \( \chi^2(2) = 21.55, p < 0.001 \); C1: \( t(33) = 4.70, p < 0.001 \); C2: \( t(33) = 4.48, p < 0.001 \)). The same group differences were found for hair nicotine levels (that is, \( \chi^2(2) = 25.40, p < 0.001 \); C1: \( t(33) = 5.44, p < 0.001 \); C2: \( t(33) = 4.77, p < 0.001 \)). These findings indicate again that infants in the IEG households were not protected from secondhand smoke exposure.

**Exploring the contribution of air, dust, surface, and finger contamination to overall exposure**

Our findings showed that infants in the IEG and DEG groups live in homes with ETS contaminated air, dust, and surfaces. To explore how air, dust, and surface contamination in living rooms and bedrooms may contribute to the overall exposure to ETS, we first examined their bivariate relations. Air and surface nicotine showed consistently positive and medium to large correlations, ranging from 0.85 (living room and bedroom surface nicotine) and 0.84 (living room and bedroom air nicotine) to 0.49 (living room air and living room surface) and 0.51 (living room surface and bedroom air). In contrast, dust nicotine levels showed low to medium correlations (<0.40) with other air and surface nicotine levels.

We examined next the extent to which air, dust, and surface nicotine levels in living rooms and bedrooms predicted average urine cotinine levels. In the subset of 27 households for which measures on all variables were available, living room and bedroom surface nicotine (\( t(21) = -2.16, p = 0.043 \); \( t(21) = 3.12, p = 0.005 \)), living room and bedroom dust nicotine (\( t(21) = -2.22, p = 0.038 \); \( t(21) = 2.07, p = 0.090 \)), and bedroom air nicotine (\( t(21) = 3.47, p = 0.002 \)) each accounted for a significant proportion of variance for a total \( R^2 = 0.78 \) (\( F(5,21) = 34.98, p < 0.001 \)).

A similar model was fit in the larger subset of 41 households for which data were available on urine cotinine, air, and surface nicotine in living rooms and bedrooms. In this sample, living room air nicotine (\( t(38) = 4.62, p < 0.001 \); semi-partial \( r^2 = 0.23 \)) and bedroom surface nicotine (\( t(38) = 2.38, p = 0.022 \); semi-partial \( r^2 = 0.06 \)) accounted for significant proportions of variance for a total \( R^2 = 0.74 \) (\( F(2,38) = 45.57, p < 0.001 \)).

**DISCUSSION**

This study investigated air, dust, surfaces, and mother’s index fingers to determine whether they are contaminated with nicotine, the single best marker of ETS and its chemical constituents. Nicotine was detected in the living and bedroom air of infants in the non-smoker and smoker households. Nicotine was also detected in dust and on surfaces of living rooms and bedrooms of infants in IEG and DEG households. Moreover, nicotine was detected on the index fingers of smoking mothers. Although IEG and DEG households had about the same amount of dust, we found three times as much nicotine per square metre in the living rooms of DEG than in IEG households, and we found about six times as much nicotine per square metre in the bedrooms of DEG and IEG households. That is, differences in amount of
of infants in IEG households were successful in reducing dust, and surface contamination and exposure levels compared to IEG households were 5–7 times higher. Average contamination of mothers' index fingers was approximately the same in the DEG and IEG households. This is consistent with the observation that mothers in the IEG and DEG groups had approximately equal smoking rates. Consistent with the different levels of contamination, infants in IEG households showed exposure levels 5–8 times higher than those of infants in NEG households. Exposure levels were 2–6 times higher in infants of DEG households than those in IEG households.

**Multiple sources of exposure**

Infants of smokers live in homes that are contaminated with ETS and are exposed to ETS. This study showed that ETS contamination is not limited to the indoor air, but includes surfaces and dust in living rooms and bedrooms and on smokers' skin. This puts infants at risk of exposure to the toxics components of ETS through multiple sources and multiple pathways, including the inhalation of contaminated air, the inhalation and ingestion of dust, ingestion and skin contact with contaminated household surfaces, and the skin of smokers.

This study provided preliminary evidence in support of the multiple exposure risk in infants. Our findings suggest that nicotine contamination of air, dust, and surfaces in living rooms and bedrooms independently account for variance in infants' urine cotinine levels. Specifically, higher levels of bedroom air, dust, and surface contamination are associated with higher levels of urine cotinine.

**Protecting infants from ETS exposure**

This study suggests that smokers can reduce household contamination and ETS exposure of their infants by implementing a strict smoking ban in the home and by not smoking in the proximity of the infant outside the home. These findings differ from those reported by Al-Delaimy et al. with respect to hair nicotine, who found no significant effect on hair nicotine levels of children (aged 3 months to 10 years) if household members smoked outside or inside the home. The fact that our sample consisted of infants under 12 months (mean 7 months) may partly explain why we found differences in exposure levels between infants in households with and without indoor smoking bans. Because Al-Delaimy et al.'s study did not include measures of ETS contamination, it is unclear whether households with and without indoor smoking bans actually differed in ETS contamination of air, dust, and surfaces. Moreover, it is unclear the extent to which ETS exposure outside the home may have contributed to the overall exposure of children in their study.

Although smoking bans appear to reduce indoor ETS contamination and ETS exposure of infants, smokers will find it difficult—if not impossible—to protect their children from ETS and its toxics components. These findings are consistent with those of Al-Delaimy et al. While parents in the IEG households were successful in reducing dust, surface, and air contamination and exposure levels compared to DEG households, they were unable to reduce ETS contamination and exposure to levels found in non-smoker households. Moreover, skin contamination did not differ between mothers in the DEG and IEG households as is expected because smoking rates were similar in the two groups.

To better understand the challenge to protect children of smokers from secondhand smoke, it is important to consider the parents’ efforts to do so. Almost 90% of parents in the IEG households always or almost always smoked outside, and approximately two thirds always or almost always closed doors and windows when smoking outside. In only four IEG households were any cigarettes reportedly smoked indoors during the study period. The average number of cigarettes reportedly smoked in the proximity of the infants in IEG households (for example, at home, in the car, or outside when child was present) was less than 0.1 per day. It appears that parents tried their best to protect their children from tobacco smoke and had reason to believe that they succeeded in doing so. While parents were able to lower ETS contamination and ETS exposure, these efforts were insufficient to achieve levels of nicotine contamination in the homes and exposure found in infants of non-smoking parents.

Our findings point to some of the sources of ETS exposure that parents cannot easily control through indoor smoking bans. ETS can remain in the home even if smoking took place days, weeks and months earlier through contaminated dust and surfaces, including the frame of an infant’s bed and a smoker’s finger. Additionally, ETS may find its way into the home through windows and doors if cigarettes are smoked outside and through contaminated clothes, skin, and dust carried into the home if cigarettes were smoked elsewhere.

This line of research has many important implications for the comprehensive measurement of ETS contamination and exposure, the study of health risks, the control of secondhand smoke, and public health policies. The comprehensive assessment of secondhand smoke contamination must consider the multiple sources of exposure, including but not limited to, air, dust, surfaces, and skin. Because ETS is not uniformly distributed throughout a home and over time, different household members may be at different risk of exposure to different sources of ETS contamination and different ETS components. For example, if exposure risks in infants are the primary concern, air samples should be taken at lower heights, and objects and surfaces should be sampled with which an infant is more likely to have contact. If smoking takes place irregularly, the duration and frequency of sampling must become an important consideration. If rooms are well ventilated during smoking, highly volatile ETS compounds and ETS particles may contribute less to long-term ETS contamination than other compounds.

Little is currently known about the differential health risks associated with the inhalation or ingestion of ETS and its toxic components or the health risks associated with ETS exposure within minutes, days, or months after tobacco smoke was emitted. As a first step, research is needed to better understand the validity of nicotine as a marker of ETS in air, dust, and surfaces over the time course of ETS contamination. Next, efforts are necessary to better measure and model the cumulative effects of exposure to ETS through different contamination sources. This and other studies suggest that dose of exposure is a complex function not only of amount of secondhand smoke, timing, and duration but also of different sources and routes of exposure.

Findings of this study suggest that interventions and public policies to reduce secondhand smoke exposure may have to be revised. The three major concerns. First, smoking outdoors, in different rooms, or when non-smokers are absent does not completely protect non-smokers from tobacco smoke, although it significantly reduces the likely level of exposure. Thus, children of smokers, non-smoking staff, and non-smokers renting or buying cars, apartments, and houses of smokers, are at risk of secondhand
smoke exposure and the associated health risks. Second, because ETS contaminates surfaces, dust, and skin, serious consideration should be given to efforts necessary to decontaminate homes, cars, furniture, etc., of smokers. Third, because contaminated indoor environments may present significant health risks to unsuspecting non-smokers, public policies may be needed, requiring disclosure of the smoking status of former tenants of apartments and offices and/or owners of cars and homes. To understand and evaluate the health risks associated with ETS exposure, we must take into account the complex physical and chemical properties of ETS, the extent and persistence of ETS contamination of residential environments, the multiple exposure pathways, the cumulative effects of ETS exposure, and the differential vulnerability of risk populations. There is yet much to be learned before we know how to comprehensively assess the risks of ETS exposure and effectively protect non-smokers from ETS.

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REFERENCES
1 California Environmental Protection Agency. Health effects of exposure to environmental tobacco smoke: final report. Sacramento, CA: The Office of Environmental Health Hazard Assessment, 1997
11 Roberts JW, Dickey P. Exposure of children to pollutants in house dust and indoor air. Rev Environ Contam Toxicol 1995;143:59–78
20 Daisey JM, Keefer L. Hair as a biomarker for passive smoking. Tobacco Control Sep 2002;11:176–82
22 Al-Delaimy WK, Crane J, Woodward A. Is the hair nicotine level a more accurate biomarker of environmental tobacco smoke exposure than urine cotinine? J Epidemiol Community Health 2002;56:66–71
24 StateCorp. State statistical software: Release 7.0. College Station, Texas: State Corporation, 2001
Households contaminated by environmental tobacco smoke: sources of infant exposures

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